

(REVIEW ARTICLE)



# A review on the molecular basis of stemness of mesenchymal stem cells

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### Abstract

Mesenchymal stem cells (MSCs) are increasingly becoming the focal point of research in regenerative medicine due to their potential to differentiate into several different lineages including those of non-mesodermal origin. With several reports of successful differentiation of MSCs into several different cell types such as osteocytes, chondrocytes, cardiocytes and even fully functional neurons, it is more important than ever to understand the stemness of these cells at a molecular level. While no stemness gene has been identified that is unique to MSCs, there have been speculations that the stemness of MSCs is maintained similarly to that of Embryonic Stem Cells (ESCs). This review takes a look at four genes, namely, NANOG, OCT4, SOX2 & KLF4, and their role in maintaining the stemness of MSCs. These genes are significant for the stemness of ESCs & therefore, could play a role in MSCs as well.

Keywords: Mesenchymal Stem Cells; NANOG; OCT4; SOX2; KLF4; Stemness; MSCs

### 1. Introduction

Mesenchymal stem cells are multipotent stem cells of mesodermal origin. They are characterized by three properties: i) cells should adhere to plastic; ii) cells should be able to differentiate into chondrocytes, osteocytes & adipocytes; iii) MSCs express cell surface markers like the cluster of differentiation CD29, CD44, CD73, CD90, CD105 and lack the expression of CD14, CD34, CD45 and HLA (human leukocyte antigen)-DR. MSCs were first discovered in the bone marrow and then several other sources such as adipose tissue, dental pulp and umbilical cord were also discovered. MSCs have been in clinical trials for treating several diseases as well as injuries such as bone restructuring, cardiovascular repair and nervous system repair. Although several studies have been conducted, very little is known about the molecular mechanism of MSC stemness. No unique transcription factor has thus far been identified that is unique to MSCs. This review will take a look at NANOG, OCT4, SOX2 & KLF4, four regulators of stemness in ESCs, and their role in MSCs [1]–[3].

### 2. Molecular Mechanism of Stemness

### 2.1. NANOG

NANOG is derived from "Tír na nÓg" of Celtic mythology which stands for "Land of the Ever Young" [4]. This is in reference to the role of the gene in maintaining the undifferentiated state of stem cells. The human NANOG gene is 305 amino acids long and contains three functional domains, namely, the homeodomain, the N-terminal domain and the C-terminal domain. NANOG is an essential transcription factor that is partly responsible for the maintenance of self-renewal and the undifferentiated state of stem cells. NANOG along with OCT4 & SOX2 is responsible for the upregulation of self-renewal and pluripotency-related genes and the downregulation of differentiation-related genes [5].

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The exact mechanism of NANOG-induced self-renewal and maintenance of an undifferentiated state is yet to be fully elucidated. There are major differences in the targets of NANOG between MSCs and ESCs which could be attributed to the difference in the epigenetics of the two cell types. NANOG regulates a unique set of genes in different cell types thus having a tissue-specific effect. This could suggest that NANOG is involved in more than just the maintenance of pluripotency and self-renewal [6].

A recent study established that the knockdown of NANOG resulted in reduced cell proliferation and induction of G0/G1 cell cycle arrest in MSCs. The paper found that NANOG knockdown resulted in the reduction in p53 expression which increased p21 & p27 levels resulting in cell cycle arrest. Also, NANOG resulted in the reduction in DNMT1 levels, which acts as a regulator of p21 & p27. NANOG knockdown also resulted in the reduction of PREF1 which is a marker for adipose mesenchymal stem cells [7].

Another study attempted to reverse the effects of ageing on MSCs by ectopic expression of NANOG. MSCs from older donors usually display reduced proliferative myogenic differentiation potential. NANOG when expressed ectopically upregulated genes of the cell cycle, DNA damage repair and replication. NANOG also significantly increased the proliferation capacity of the cells. This is especially helpful for elderly patients that suffer from diseases that can be treated with MSC therapy [8]. Ectopic expression of NANOG also improved the contractile function of senescent MSCs in three-dimensional microtissues. NANOG reversed the effects of stem cell senescence as well as restored the pluripotency and self-renewal capacity of senescent MSCs [9]. The molecular basis of restoring the self-renewal capacity of senescent MSCs [9]. NANOG upregulates PBX1 which in turn increases the activity of the PI3K/AKT signalling pathway. The PI3K/AKT signalling pathway regulates cell proliferation, DNA repair & senescence [10].

# 2.2. OCT4

OCT4 is an octamer-binding transcription factor encoded in humans by the gene POU5F1. It consists of an octamer motif which binds for activating or deactivating the target genes [11].

There are conflicting reports about whether OCT4 is expressed in MSCs or not. Some studies suggest that it is expressed, albeit in low concentrations [12], [13], while others claim that it is not expressed [14], [15].

Regardless, ectopic expression of OCT4 results in the enhanced proliferation of MSCs as was seen in a study conducted in 2014. The cells saw an increase in the levels of cyclins that are responsible for cell proliferation [16]. Another study found that ectopic expression of OCT4 results in the upregulation of NANOG & SOX2. This could explain the molecular mechanism behind the increased proliferative & differentiation capacity of the cells [17].

Similar to NANOG which affects p21 through DNMT1, ectopically expressed OCT4 can also improve the self-renewal capacity of MSCs through DNMTs. OCT4 upregulated DNMTs which in turn methylated the promoters of p21 resulting in its inhibition. This resulted in the reversal of cell senescence and restored its self-renewal capacity [18].  $\beta$ -catenin has been found to regulate OCT4. A 2022 paper established that  $\beta$ -catenin increased the survival of MSCs and promoted angiogenesis in the case of stem cell therapy for cardiac diseases.  $\beta$ -catenin positively regulates OCT4 which in turn regulates genes such as Ang1, bFGF, HGF, VEGF, Bcl2, and surviving which are responsible for the survival and angiogenesis of MSCs [19].

# 2.3. SOX2

SOX2 belongs to the highly conserved Sox family of transcription factors that play key roles in mammalian development. The Sox family was first discovered in 1990 during the discovery of the mammalian testis-determining factor 'Sry' [20]. SOX2 is the third element in the core transcription factors that govern the pluripotency of ESCs. SOX2 is activated by KLF4 (another transcription factor) as a result of the JAK-STAT signalling pathway [21]. SOX2 binds to several regions in the DNA using the HMG domain at the minor groove. This is in juxtaposition from OCT4 which binds to the major groove [22], [23].

Interestingly, the levels of SOX2 need to be tightly regulated as high or low levels of SOX2 can induce differentiation in the stem cells [22], [24]. Knockdown studies have established that reduced expression of SOX2 in hESCs results in loss of the undifferentiated state of the stem cell, changed cell morphology and cell marker expression as well as an increase in the expression of trophectoderm markers. It also, notably, results in a reduction in the expressions of NANOG & OCT4 thus again establishing the interconnected nature of the three master transcription factors [25]. SOX2 forms a heterodimer with OCT4 to bind to several sites in the DNA for the regulation of numerous genes [26]. Apart from

working in tandem with OCT4, some studies have established that SOX2, while being dispensable for the activation of the Oct-Sox heterodimer enhancer activation, is a key component in the regulation of OCT4 [27].

SOX2 expression in MSCs has been confirmed by several studies [28]. The overexpression of SOX2 in MSCs results in enhanced cell proliferation as a result of increased levels of cyclin D1 [16]. Interestingly, the expression of SOX2 is of great importance in the case of low-density MSC culturing & knockdown of the gene resulting in the inhibition of multipotentiality and cell proliferation [29]. One method of maintaining the self-renewal capacity of MSCs is through YAP1 which is directly regulated by SOX2 [30]. Also, SOX2 regulates DKK1 as well as c-Myc which determines the lineage differentiation of MSCs [31].

Apart from maintaining the pluripotency of the MSCs, SOX2 can also result in the differentiation of the MSCs into neuronlike cells [32]. A recent study used plant phenol Resveratrol on MSCs and established its capacity to maintain the pluripotency of the MSCs in vitro. On a molecular level, the compound acts as a SIRT1 activator which in turn maintains the levels of SOX2 required for pluripotency in the MSCs for extended culture durations. This could have great utility in MSC therapy for bone regeneration and other regenerative therapies that require extensive culturing [33]. Another interesting study established that apoptotic vesicles from ESCs could promote skin wound healing by transferring SOX2 to skin MSCs via the activation of the Hippo signalling pathway. This could open new avenues in regeneration therapy [34].

### 2.4. KLF4

KLF4 belongs to the family of SP1-like transcription factors [35], [36]. KLF4 is one of the four factors that was identified as part of the factors required for reprogramming fibroblasts into induced pluripotent stem cells (iPSCs) [37]. KLF4 is an important transcription factor in maintaining the pluripotency and differentiation capacities of ESCs and it does so through several pathways. One of these pathways is the activation of Lefty1 with OCT4 & SOX2 [38]. Also, the overexpression of KLF4 along with PBX1 upregulates NANOG in ESCs which results in the maintenance of the undifferentiated state of the cells [39].

The knockdown of KLF4 results in the commitment of ESCs to the endodermal lineage. This is because KLF4 is responsible for negatively regulating the endodermal marker expression [40]. The knockdown studies have been further validated by other studies. In a study conducted on the function of ERK1 & ERK2, the two molecules phosphorylated KLF4 which resulted in its inactivation which ultimately induced embryonic stem cell differentiation [41]. Loss of KLF4 in mice resulted in altered differentiation & proliferation as well as precancerous changes in the adult stomach [42]. This shows that the function of KLF4 isn't just restricted to stem cells.

The importance of KLF4 for the maintenance of the undifferentiated state of MSCs has been established by several studies [43]. Many recent studies focus on the repression of KLF4 in MSCs. A 2023 study established that CUL4B promotes mesenchymal stem cell commitment to osteogenic lineage by repressing KLF4 [44]. Another repressor of KLF4 a miRNA called miR-10a. This miRNA reduces with ageing in MSCs. Upregulation of miR-10a resulted in increased adipogenic, osteogenic and, chondrogenic differentiation & reduced senescence while downregulation resulted in increased cell senescence and decreased differentiation potential. This happens as a result of miR-10a binding and suppressing the expression of KLF4 [45]. These old MSCs with overexpressed miR-10a when implanted into infarcted mouse hearts displayed better stem cell survival, improved cardiac function & increased angiogenesis in comparison to the control [46]. MSCs-derived extracellular vesicles have been used to treat several diseases one of which is rheumatoid arthritis. MSC-derived EVs containing miR-21 targeting Tet1 which upregulates KLF4 was established to be a successful treatment for rheumatoid arthritis in mouse models [47].

# 3. Conclusion

Even though studies have shown that the ectopic expression of NANOG, SOX2, & OCT4 have maintained the pluripotency and the self-renewal capacity of MSCs in culture, and the repression of KLF4 has resulted in loss of pluripotency, the expression of these genes under normal conditions is highly debated. With conflicting reports proving their expression as well as inexpression, more robust studies need to be conducted to provide an irrefutable consensus. Also, further studies need to be conducted to elucidate the exact molecular mechanism of the maintenance of pluripotency and self-renewal of these cells.

#### **Compliance with ethical standards**

#### Disclosure of conflict of interest

No conflict of interest to be disclosed.

#### References

- T. M. Liu\*, 'Stemness of Mesenchymal Stem Cells', J. Stem Cell Ther. Transplant., vol. 1, no. 1, pp. 071–073, Dec. 2017.
- [2] I. Ullah, R. B. Subbarao, and G. J. Rho, 'Human mesenchymal stem cells current trends and future prospective', Biosci. Rep., vol. 35, no. 2, p. e00191, Apr. 2015, doi: 10.1042/BSR20150025.
- [3] Y. Han, X. Li, Y. Zhang, Y. Han, F. Chang, and J. Ding, 'Mesenchymal Stem Cells for Regenerative Medicine', Cells, vol. 8, no. 8, Art. no. 8, Aug. 2019, doi: 10.3390/cells8080886.
- [4] C. I et al., 'Functional expression cloning of NANOG, a pluripotency sustaining factor in embryonic stem cells', Cell, vol. 113, no. 5, May 2003, doi: 10.1016/s0092- 8674(03)00392-1.
- [5] N. Gawlik-Rzemieniewska and I. Bednarek, 'The role of NANOG transcriptional factor in the development of malignant phenotype of cancer cells', Cancer Biol. Ther., vol. 17, no. 1, pp. 1–10, Nov. 2015, doi: 10.1080/15384047.2015.1121348.
- [6] T. M. Liu, Y. N. Wu, X. M. Guo, J. H. P. Hui, E. H. Lee, and B. Lim, 'Effects of ectopic NANOG and OCT4 overexpression on mesenchymal stem cells', Stem Cells Dev., vol. 18, no. 7, pp. 1013–1022, Sep. 2009, doi: 10.1089/scd.2008.0335.
- [7] M. Pitrone et al., 'Knockdown of NANOG Reduces Cell Proliferation and Induces G0/G1 Cell Cycle Arrest in Human Adipose Stem Cells', Int. J. Mol. Sci., vol. 20, no. 10, Art. no. 10, Jan. 2019, doi: 10.3390/ijms20102580.
- [8] J. Han, P. Mistriotis, P. Lei, D. Wang, S. Liu, and S. T. Andreadis, 'NANOG Reverses the Effects of Organismal Aging on Mesenchymal Stem Cell Proliferation and Myogenic Differentiation Potential', Stem Cells, vol. 30, no. 12, pp. 2746–2759, Dec. 2012, doi: 10.1002/stem.1223.
- [9] A. Shahini, P. Mistriotis, M. Asmani, R. Zhao, and S. T. Andreadis, 'NANOG Restores Contractility of Mesenchymal Stem Cell-Based Senescent Microtissues', Tissue Eng. Part A, vol. 23, no. 11–12, pp. 535–545, Jun. 2017, doi: 10.1089/ten.tea.2016.0494.
- [10] F. Liu et al., 'NANOG Attenuates Hair Follicle-Derived Mesenchymal Stem Cell Senescence by Upregulating PBX1 and Activating AKT Signaling', Oxid. Med. Cell. Longev., vol. 2019, p. 4286213, Dec. 2019, doi: 10.1155/2019/4286213.
- [11] J. Takeda, S. Seino, and G. I. Bell, 'Human Oct3 gene family: cDNA sequences, alternative splicing, gene organization, chromosomal location, and expression at low levels in adult tissues.', Nucleic Acids Res., vol. 20, no. 17, pp. 4613–4620, Sep. 1992.
- [12] U. Riekstina et al., 'Embryonic Stem Cell Marker Expression Pattern in Human Mesenchymal Stem Cells Derived from Bone Marrow, Adipose Tissue, Heart and Dermis', Stem Cell Rev. Rep., vol. 5, no. 4, pp. 378–386, Dec. 2009, doi: 10.1007/s12015-009-9094-9.
- [13] T. Tondreau et al., 'Mesenchymal Stem Cells Derived from CD133-Positive Cells in Mobilized Peripheral Blood and Cord Blood: Proliferation, OCT4 Expression, and Plasticity', Stem Cells, vol. 23, no. 8, pp. 1105–1112, Sep. 2005, doi: 10.1634/stemcells.2004-0330.
- [14] T. Mueller, J. Luetzkendorf, K. Nerger, H.-J. Schmoll, and L. P. Mueller, 'Analysis of OCT4 expression in an extended panel of human tumor cell lines from multiple entities and in human mesenchymal stem cells', Cell. Mol. Life Sci., vol. 66, no. 3, p. 495, Nov. 2008, doi: 10.1007/s00018-008-8623-z.
- [15] E. Pierantozzi et al., 'Pluripotency Regulators in Human Mesenchymal Stem Cells: Expression of NANOG But Not of OCT-4 and SOX-2', Stem Cells Dev., vol. 20, no. 5, pp. 915–923, May 2011, doi: 10.1089/scd.2010.0353.
- [16] S.-M. Han et al., 'Enhanced proliferation and differentiation of OCT4- and SOX2- overexpressing human adipose tissue mesenchymal stem cells', Exp. Mol. Med., vol. 46, no. 6, Art. no. 6, Jun. 2014, doi: 10.1038/emm.2014.28.

- [17] K.-H. Wang, A.-P. Kao, C.-C. Chang, T.-C. Lin, and T.-C. Kuo, 'Upregulation of NANOG and Sox-2 genes following ectopic expression of Oct-4 in amniotic fluid mesenchymal stem cells', Biotechnol. Appl. Biochem., vol. 62, no. 5, pp. 591–597, 2015, doi: 10.1002/bab.1315.
- [18] Y. Lu et al., 'OCT4 maintains self-renewal and reverses senescence in human hair follicle mesenchymal stem cells through the downregulation of p21 by DNA methyltransferases', Stem Cell Res. Ther., vol. 10, no. 1, p. 28, Jan. 2019, doi: 10.1186/s13287-018-1120-x.
- [19] P. Wang et al., 'β-Catenin promotes long-term survival and angiogenesis of peripheral blood mesenchymal stem cells via the OCT4 signaling pathway', Exp. Mol. Med., vol. 54, no. 9, Art. no. 9, Sep. 2022, doi: 10.1038/s12276-022-00839-4.
- [20] A. H. Sinclair et al., 'A gene from the human sex-determining region encodes a protein with homology to a conserved DNA-binding motif', Nature, vol. 346, no. 6281, pp. 240–244, Jul. 1990, doi: 10.1038/346240a0.
- [21] H. Niwa, K. Ogawa, D. Shimosato, and K. Adachi, 'A parallel circuit of LIF signalling pathways maintains pluripotency of mouse ES cells', Nature, vol. 460, no. 7251, pp. 118–122, Jul. 2009, doi: 10.1038/nature08113.
- [22] S. Zhang and W. Cui, 'SOX2, a key factor in the regulation of pluripotency and neural differentiation', World J. Stem Cells, vol. 6, no. 3, pp. 305–311, Jul. 2014, doi: 10.4252/wjsc.v6.i3.305.
- [23] I. Chambers and S. R. Tomlinson, 'The transcriptional foundation of pluripotency', Dev. Camb. Engl., vol. 136, no. 14, pp. 2311–2322, Jul. 2009, doi: 10.1242/dev.024398.
- [24] J. L. Kopp, B. D. Ormsbee, M. Desler, and A. Rizzino, 'Small increases in the level of SOX2 trigger the differentiation of mouse embryonic stem cells', Stem Cells Dayt. Ohio, vol. 26, no. 4, pp. 903–911, Apr. 2008, doi: 10.1634/stemcells.2007-0951.
- [25] H. Fong, K. A. Hohenstein, and P. J. Donovan, 'Regulation of self-renewal and pluripotency by SOX2 in human embryonic stem cells', Stem Cells Dayt. Ohio, vol. 26, no. 8, pp. 1931–1938, Aug. 2008, doi: 10.1634/stemcells.2007-1002.
- [26] N. Tapia et al., 'Dissecting the role of distinct OCT4-SOX2 heterodimer configurations in pluripotency', Sci. Rep., vol. 5, no. 1, Art. no. 1, Aug. 2015, doi: 10.1038/srep13533.
- [27] S. Masui et al., 'Pluripotency governed by SOX2 via regulation of Oct3/4 expression in mouse embryonic stem cells', Nat. Cell Biol., vol. 9, no. 6, pp. 625–635, Jun. 2007, doi: 10.1038/ncb1589.
- [28] P. Gil-Kulik et al., 'Evaluation of the Impact of Pregnancy-Associated Factors on the Quality of Wharton's Jelly-Derived Stem Cells Using SOX2 Gene Expression as a Marker', Int. J. Mol. Sci., vol. 23, no. 14, Art. no. 14, Jan. 2022, doi: 10.3390/ijms23147630.
- [29] D. S. Yoon, Y. H. Kim, H. S. Jung, S. Paik, and J. W. Lee, 'Importance of SOX2 in maintenance of cell proliferation and multipotency of mesenchymal stem cells in low- density culture', Cell Prolif., vol. 44, no. 5, pp. 428–440, 2011, doi: 10.1111/j.1365-2184.2011.00770.x.
- [30] E. Seo et al., 'SOX2 Regulates YAP1 to Maintain Stemness and Determine Cell Fate in the Osteo-Adipo Lineage', Cell Rep., vol. 3, no. 6, pp. 2075–2087, Jun. 2013, doi: 10.1016/j.celrep.2013.05.029.
- [31] S. B. Park et al., 'SOX2 has a crucial role in the lineage determination and proliferation of mesenchymal stem cells through Dickkopf-1 and c-MYC', Cell Death Differ., vol. 19, no. 3, pp. 534–545, Mar. 2012, doi: 10.1038/cdd.2011.137.
- [32] Y. Qin, C. Zhou, N. Wang, H. Yang, and W.-Q. Gao, 'Conversion of Adipose Tissue- Derived Mesenchymal Stem Cells to Neural Stem Cell-Like Cells by a Single Transcription Factor, SOX2', Cell. Reprogramming, vol. 17, no. 3, pp. 221–226, Jun. 2015, doi: 10.1089/cell.2015.0001.
- [33] Y. Choi et al., 'Enhancement of Mesenchymal Stem Cell-Driven Bone Regeneration by Resveratrol-Mediated SOX2 Regulation', Aging Dis., vol. 10, no. 4, pp. 818–833, Aug. 2019, doi: 10.14336/AD.2018.0802.
- [34] Y. Qu et al., 'Apoptotic vesicles inherit SOX2 from pluripotent stem cells to accelerate wound healing by energizing mesenchymal stem cells', Acta Biomater., vol. 149, pp. 258–272, Sep. 2022, doi: 10.1016/j.actbio.2022.07.009.
- [35] A. R. Black, J. D. Black, and J. Azizkhan-Clifford, 'Sp1 and krüppel-like factor family of transcription factors in cell growth regulation and cancer', J. Cell. Physiol., vol. 188, no. 2, pp. 143–160, 2001, doi: 10.1002/jcp.1111.
- [36] J. M. Shields, R. J. Christy, and V. W. Yang, 'Identification and Characterization of a Gene Encoding a Gut-enriched Krüppel-like Factor Expressed during Growth Arrest', J. Biol. Chem., vol. 271, no. 33, pp. 20009–20017, Aug. 1996.

- [37] K. Takahashi and S. Yamanaka, 'Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors', Cell, vol. 126, no. 4, pp. 663–676, Aug. 2006, doi: 10.1016/j.cell.2006.07.024.
- [38] Y. Nakatake et al., 'KLF4 Cooperates with Oct3/4 and SOX2 To Activate the Lefty1 Core Promoter in Embryonic Stem Cells', Mol. Cell. Biol., vol. 26, no. 20, pp. 7772–7782, Oct. 2006, doi: 10.1128/MCB.00468-06.
- [39] K. K.-K. Chan et al., 'KLF4 and PBX1 Directly Regulate NANOG Expression in Human Embryonic Stem Cells', Stem Cells, vol. 27, no. 9, pp. 2114–2125, Sep. 2009, doi: 10.1002/stem.143.
- [40] I. Aksoy et al., 'KLF4 and Klf5 differentially inhibit mesoderm and endoderm differentiation in embryonic stem cells', Nat. Commun., vol. 5, no. 1, Art. no. 1, Apr. 2014, doi: 10.1038/ncomms4719.
- [41] M. O. Kim et al., 'ERK1 and ERK2 regulate embryonic stem cell self-renewal through phosphorylation of KLF4', Nat. Struct. Mol. Biol., vol. 19, no. 3, Art. no. 3, Mar. 2012, doi: 10.1038/nsmb.2217.
- [42] J. P. Katz et al., 'Loss of KLF4 in mice causes altered proliferation and differentiation and precancerous changes in the adult stomach', Gastroenterology, vol. 128, no. 4, pp. 935–945, Apr. 2005, doi: 10.1053/j.gastro.2005.02.022.
- [43] N. Saulnier et al., 'Gene profiling of bone marrow- and adipose tissue-derived stromal cells: a key role of Kruppellike factor 4 in cell fate regulation', Cytotherapy, vol. 13, no. 3, pp. 329–340, Mar. 2011, doi: 10.3109/14653249.2010.515576.
- [44] R. Yu et al., 'CUL4B orchestrates mesenchymal stem cell commitment by epigenetically repressing KLF4 and C/EBPδ', Bone Res., vol. 11, no. 1, Art. no. 1, Jun. 2023, doi: 10.1038/s41413-023-00263-y.
- [45] J. Li et al., 'miR-10a restores human mesenchymal stem cell differentiation by repressing KLF4', J. Cell. Physiol., vol. 228, no. 12, pp. 2324–2336, 2013, doi: 10.1002/jcp.24402.
- [46] J. Dong et al., 'miR-10a rejuvenates aged human mesenchymal stem cells and improves heart function after myocardial infarction through KLF4', Stem Cell Res. Ther., vol. 9, no. 1, p. 151, May 2018, doi: 10.1186/s13287-018-0895-0.
- [47] G.-Q. Li et al., 'MicroRNA-21 from bone marrow mesenchymal stem cell-derived extracellular vesicles targets TET1 to suppress KLF4 and alleviate rheumatoid arthritis', Ther. Adv. Chronic Dis., vol. 12, p. 20406223211007370, Jan. 2021, doi: 10.1177/20406223211007369.